

Gene network reconstruction from microarray data
 F. Jaffezeic (1) and G. Tosser-Klopp (2)
 1 INRA, Jouy-en-Josas, France
 2 INRA, Toulouse, France

<p>Gene network reconstruction from microarray data</p>  <h2>Gene network reconstruction from microarray data</h2> <p>F. Jaffrézic¹ and G. Tosser-Klopp²</p> <p>¹INRA, Jouy-en-Josas, France - ² INRA, Toulouse, France</p> <p>Eadgene workshop, November 2008</p> 	<p>Gene network reconstruction from microarray data</p> <h2>Outline</h2> <ol style="list-style-type: none"> 1 Introduction 2 Graphical Gaussian models 3 Application
<p>Gene network reconstruction from microarray data</p> <h2>Outline</h2> <ol style="list-style-type: none"> 1 Introduction 2 Graphical Gaussian models 3 Application 	<p>Gene network reconstruction from microarray data</p> <h2>Introduction</h2> <p>Gene network reconstruction from microarray data.</p> <p>Two main approaches :</p> <ol style="list-style-type: none"> 1) Bayesian networks. 2) Graphical Gaussian models.
<p>Gene network reconstruction from microarray data</p> <h2>Introduction</h2> <p>Bayesian networks :</p> <ul style="list-style-type: none"> • Directed acyclic graph. <p>⇒ No feedback loop available</p> <ul style="list-style-type: none"> • Very computationally intensive. • No R package available ? 	<p>Gene network reconstruction from microarray data</p> <h2>Introduction</h2> <p>Graphical Gaussian models :</p> <ul style="list-style-type: none"> • Undirected graphs. • Very computationally efficient. • R package GeneNet (K. Strimmer) Schäfer and Strimmer (2005).

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<p>Gene network reconstruction from microarray data</p> <p>Introduction</p> <p>Graphical Gaussian models</p> <p>Application</p> <h2 style="text-align: center;">Introduction</h2> <p>Wehrli et al. (<i>Bioinformatics</i>, 2006) : Comparison study between Bayesian Networks and Graphical Gaussian models.</p> <p>Both methods were found to have quite similar performances for gene network reconstruction from microarray data.</p> <p>For simplicity of implementation \Rightarrow Graphical Gaussian models.</p>	<p>Gene network reconstruction from microarray data</p> <p>Introduction</p> <p>Graphical Gaussian models</p> <p>Application</p> <h2 style="text-align: center;">Outline</h2> <ol style="list-style-type: none"> 1 Introduction 2 Graphical Gaussian models 3 Application
<p>Gene network reconstruction from microarray data</p> <p>Introduction</p> <p>Graphical Gaussian models</p> <p>Application</p> <h2 style="text-align: center;">Graphical Gaussian models</h2> <p>Let \mathbf{X} be the observed data matrix with N rows (samples) and G columns (genes).</p> <p>\mathbf{X} is supposed to be drawn from a multivariate normal distribution $\mathcal{N}_G(\boldsymbol{\mu}, \boldsymbol{\Sigma})$, with mean vector $\boldsymbol{\mu} = (\mu_1, \dots, \mu_G)'$ and positive-definite covariance matrix $\boldsymbol{\Sigma} = (\sigma_{ij})_{(1 \leq i, j, \leq G)}$.</p>	<p>Gene network reconstruction from microarray data</p> <p>Introduction</p> <p>Graphical Gaussian models</p> <p>Application</p> <h2 style="text-align: center;">Graphical Gaussian models</h2> <p>Covariance parameters σ_{ij} can also be written as :</p> $\sigma_{ij} = \rho_{ij}\sigma_i\sigma_j$ <p>where σ_i^2 and σ_j^2 are the variance terms for genes i and j, respectively and ρ_{ij} is the Pearson correlation coefficient between genes i and j.</p>
<p>Gene network reconstruction from microarray data</p> <p>Introduction</p> <p>Graphical Gaussian models</p> <p>Application</p> <h2 style="text-align: center;">Graphical Gaussian models</h2> <p>Let \mathbf{P} be the Pearson correlation matrix $\mathbf{P} = (\rho_{ij})_{(1 \leq i, j, \leq G)}$.</p> <p>A high correlation coefficient ρ_{ij} between two genes may indicate either :</p> <ol style="list-style-type: none"> 1) a direct interaction between genes i and j. 2) an indirect interaction between these two genes. 3) a regulation of the two genes by a common gene. 	<p>Gene network reconstruction from microarray data</p> <p>Introduction</p> <p>Graphical Gaussian models</p> <p>Application</p> <h2 style="text-align: center;">Graphical Gaussian models</h2> <p>For network reconstruction, we are only interested in direct interactions, represented by the partial correlation matrix $\boldsymbol{\Pi} = (\pi_{ij})$.</p> <p>Coefficient π_{ij} represents the correlation between two genes i and j conditioned on all the other genes.</p>

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Graphical Gaussian models

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It can be shown that **partial correlation matrix Π** is related to the **inverse of the covariance matrix Σ** as follows :

$$\pi_{ij} = -\omega_{ij} / \sqrt{\omega_{ii}\omega_{jj}}$$

with $\Sigma^{-1} = (\omega_{ij})$, for $1 \leq i, j \leq G$.



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Construction of a GGM network.

- 1) **Estimate the empirical covariance matrix**
 $\hat{S} = (\hat{\sigma}_{ij}) = \frac{1}{N-1}(\mathbf{X} - \bar{\mathbf{X}})(\mathbf{X} - \bar{\mathbf{X}})$.
- 2) Estimate the **partial correlation matrix** using the previous equations.
- 3) **Statistical tests** to determine the **partial correlation coefficients significantly different from 0**, which correspond to the significant edges of the graph.



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Procedure **only applicable when sample size N larger than the number of variables G** , otherwise the sample covariance matrix is not positive-definite and cannot be inverted, which prevents a direct computation of the partial correlation matrix.

In microarray analyses, $N \ll G$ and small sample size N .



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Shrunk estimate of the covariance matrix using a **James-Stein estimator** (Schäfer and Strimmer, 2005).

Objectives :

- 1) Construct a **positive-definite matrix** (all eigenvalues > 0).
 - 2) **Well-conditioned covariance matrix** (ratio of its maximum and minimum singular value is not too large).
- \Rightarrow The matrix has full rank and can easily be inverted.



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Shrinkage estimation of the covariance matrix.

Let λ be a **shrinkage coefficient** ($\lambda \in [0, 1]$)

$\Sigma^* = \lambda \mathbf{T} + (1 - \lambda)\hat{S}$ where \hat{S} is the estimated empirical covariance matrix.

Shrinkage parameter λ : chosen to minimize the mean-squared error (MSE) and determined analytically (Ledoit and Wolf, 2003).



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Choice for matrix \mathbf{T} : Several possibilities.

Schäfer and Strimmer (2005) recommend for gene network reconstruction to **shrink the correlation terms towards zero** and to **leave the diagonal terms as estimated by the empirical variances**.

In this case, the **shrinkage parameter λ** can be estimated analytically as :

$$\hat{\lambda} = \frac{\sum_{i \neq j} \widehat{\text{Var}}(s_{ij})}{\sum_{i \neq j} s_{ij}^2}$$

where s_{ij} are the empirical covariance parameters.



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Test of the significant edges.

Network structure is usually **sparse**.

An **edge-specific local FDR** was defined using the estimated **partial correlation coefficients** (Schäfer and Strimmer, 2005).

An edge is considered significant if its **local FDR is smaller than 0.2** (Efron, 2005).



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Chicken infection data set

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GeneNet package.

1. List of DE genes between MM8 and MA8 :

85 differentially expressed genes at a 5% BH threshold.

No missing values allowed in GeneNet \Rightarrow **58 genes**.



Application

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Network reconstruction using the expression values for each condition independently.

No significant edges at a 20% local FDR threshold for either **MM8** or **MA8** expression values.



Application

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2. List of DE genes between MM8 and MM24 :

116 differentially expressed genes at a 1% BH threshold.

No missing values \Rightarrow **85 genes**.



Application

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Network reconstruction using the expression values for each condition independently.

For MM8 :

2356 significant edges among the 85 genes at a **20% local FDR threshold**.

1964 significant edges at a **5% local FDR threshold**.



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For MM24 :

1760 significant edges among the 85 genes at a 20% local FDR threshold.

1156 significant edges at a 5% local FDR threshold.

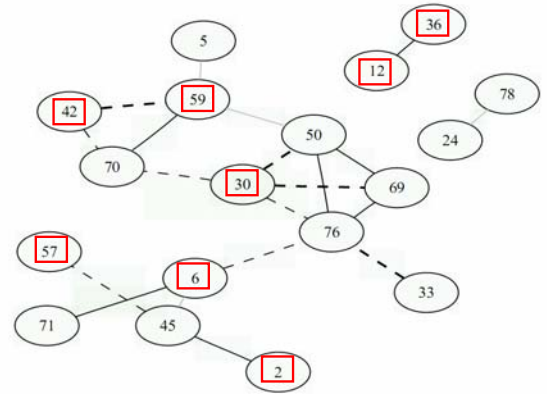
Graph of the 20 most significant edges for both conditions.



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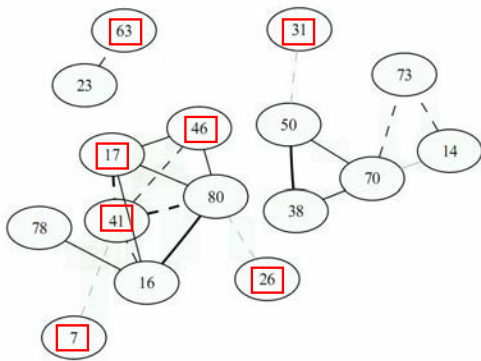
Gene network for condition MM8



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Gene network for condition MM24



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Very little overlapping between the two networks.

Only genes 50, 70 and 78 remain in both graphs.

No link in common between the two graphs.

⇒ Evolution of the gene connections over time.



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Biological interpretation

In collaboration with **Gwenola Tosser-Klopp** (INRA, Toulouse).

For condition MM8 :

Among the 18 genes present in the graph only 8 were annotated.

For condition MM24 :

Among the 16 genes present in the graph only 7 were annotated.



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Biological interpretation

For both graphs, the links found here between annotated genes were not confirmed with either Ingenuity or Pathway Studio.

⇒ Further biological validation required (using knock-out experiments, etc.).



